

Vitamin D:

How good are our assays?

Ronda Greaves



Overview

- Background
- Clinical
- Reference intervals
- Measurement systems
- Approaches to quality

The Vitamin Alphabet

- A - Retinol
- B - group of 8
- C - Ascorbic acid
- **D** - **Ergocalciferol**
- **D** - **Cholecalciferol**
- E - Tocopherol
- K - Phylloquinone

B1	Thiamine
B2	Riboflavin
B3	Niacin
(B4)	Adenine)
B5	Pantothenic acid
B6	Pyridoxine
B7 (H)	Biotin
(B8)	Inositol)
B9	Folate
(B10)	PABA)
(B11)	Choline)
B12	Cobalamin

Total: 13 = 4 fat soluble + 9 water soluble

() indicates B group compound no longer classified as vitamins

Vitamin D: Definitions

VITAMIN

An organic compound required as a nutrient, which cannot be synthesized in adequate amounts, and therefore must be obtained in the diet

HORMONE

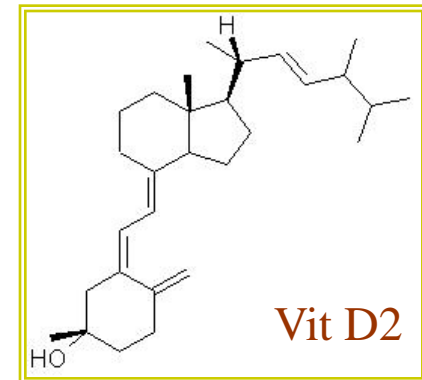
A chemical secreted by a group of cells (gland) into the circulation to affect the function of cells, through interaction with their receptors, in another part of the body

Vitamin D:

- **Vit D1** - is a 1:1 mixture of lumisterol and vitamin D2.

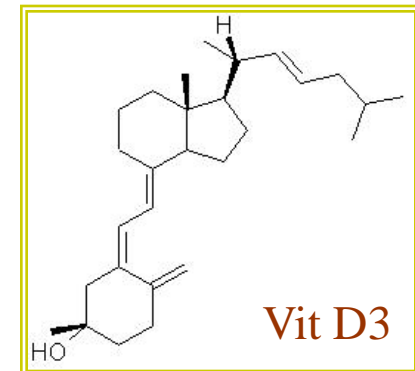
- **Vit D2 – ERGOCALCIFEROL**

- n Plant origin
- n Arises from ultraviolet irradiation of ergosterol
- n Cleaved at the 9,10 bond & develops a double bond b/w C-10 & 19

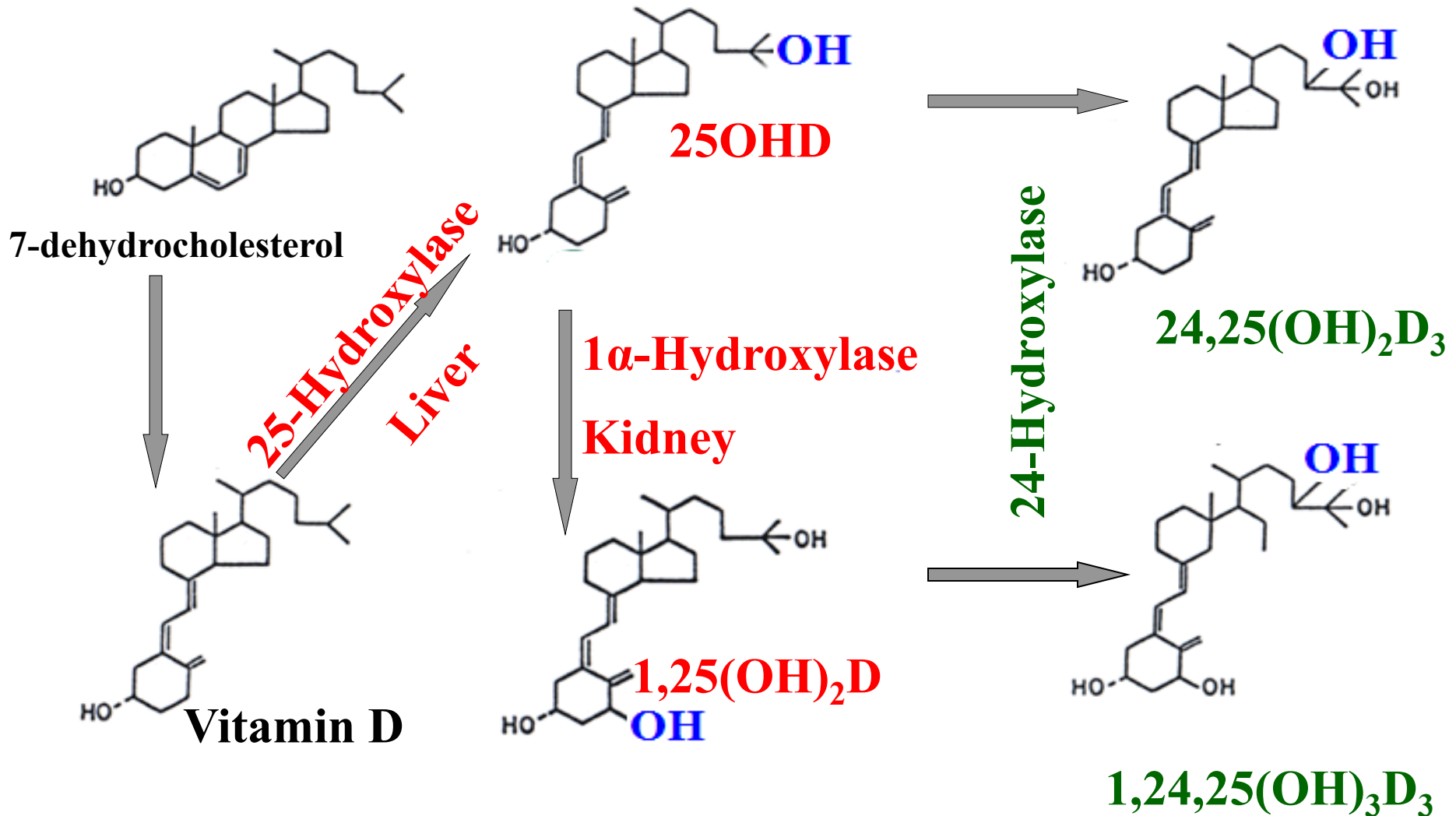


- **Vit D3 – CHOLECALCIFEROL**

- n Animal origin
- n Formed by breakage of the 9,10 bond in 7-dehydrocholesterol by ultraviolet irradiation, yielding a double bond b/w C-10 and C-19
- n Found in the skin, fur, and feathers of animals and birds exposed to sunlight, and also in butter, brain, fish oils, and egg yolk



Vitamin D



Clinical Utility

- Classically
 - n Rickets
 - n Osteomalacia
- Modern era
 - n Bone health
 - n Diabetes
 - n Autoimmune diseases
 - n Immune regulation
 - n Infections
 - n Cancer
 - n Cardiovascular disease



Before vitamin D treatment



After 14 months of vitamin D treatment

(b)

Photos from Lehninger, Principles of Biochemistry



Increasing testing numbers

“In 2009, US laboratories were reporting surges in the number of vitamin D tests being ordered - increases of 50% to even 100%. But beyond the growth in testing and usage, what's the quality required by this type of testing?”

www.Westgard.com

Vitamin D: What level is appropriate?

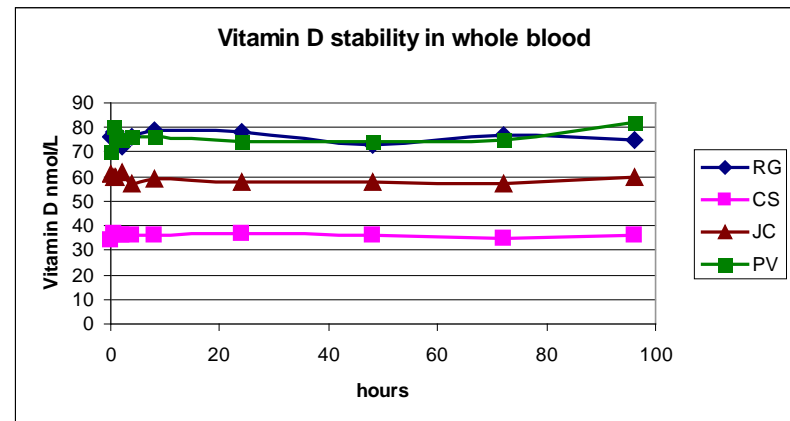
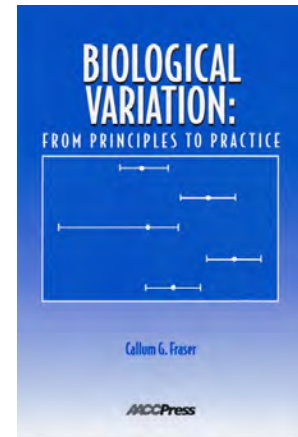
- RCH
 - n 1990's: Reference range quoted 23 to 90 nmol/L
 - n 2000's: Change to recommended range of 50 to 150 nmol/L
- Other ranges
 - n >60 nmol/L proposed based on rise in PTH
 - n >75 nmol/L proposed for health
 - n >100 nmol/L for cancer prevention
- On going debate of what range is needed for health
- **BUT** – we don't have harmonisation of methods!!!!

Pre analytical factors

- Biological Variation
- Seasonal variation
- Skin pigmentation
- Racial differences

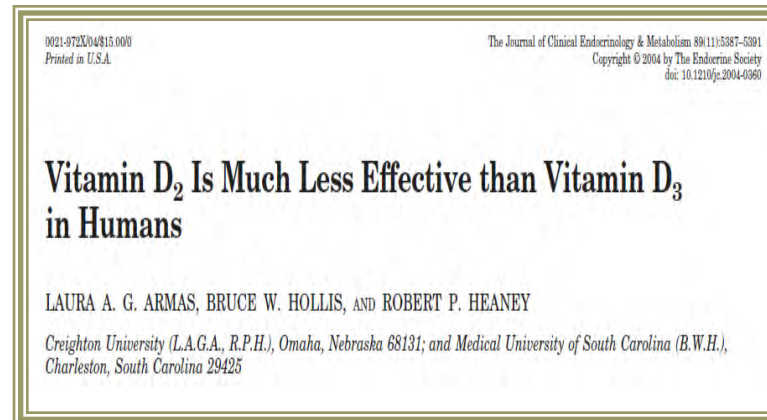
- Vitamin D is stable in whole blood stored at room temperature in sunlight for up to 96 hours.

(Poster: AACB ASM in 2005)



Vitamin D

- Choice for routine assessment of vitamin D status
- Need to extract to remove Vit D from DBP (Vit D binding protein)
- Standards calibrated against D3
- Some immunoassays cross react with D2
 - Traditionally considered an advantage
- Supplementation in Australia originally D2 now mainly D3



Vitamin D: Automated analysis

- Roche Cobas e601

- n 25 OH Vit D3 only
- n 0% cross reactivity with 25 OH Vit D2



- Diasorin Liasion

- n 25 OH Vit D3
- n >80% cross reactivity with 25 OH Vit D2



Vitamin D: Other Immunoassays

- Enzyme Linked Immunosorbent Assay
- Radio-immunoassay
 - n Diasorin (Sorin)
 - n IDS
- NEW AUTOMATED
 - n IDS – ISYS platform
 - n Abbott – recent lab trials
 - n Siemens – under development



IDS-ISYS 25-Hydroxy Vitamin D Assay

IDS-ISYS 25-Hydroxy Vitamin D Assay 100 Tests

Product Code
IS-2700

Features and Benefits

- Fully Automated Chemiluminescent Method on the IDS-ISYS System
- Wide reportable range: 12.5 - 350 nmol/L
- Analytical Sensitivity is 4.3 nmol/L
- Functional Sensitivity is 13.8 nmol/L

Product Description

The IDS-ISYS 25-Hydroxy Vitamin D assay is intended for the quantitative determination of 25-hydroxyvitamin D and other hydroxylated metabolites in human serum or plasma on the IDS-ISYS Automated Analyzer.

Related Documents

Chromatography + MS (+MS)

- Gold standard
- TAT a problem
- Expertise required
- Up front cost high



Live rates at 2010.05.23 11:09:07 UTC

500,000.00 AUD = 7,896,262,319.09 VND

Australia Dollars Vietnam Dong

1 AUD = 15,792.52 VND 1 VND = 0.0000633211 AUD

THE ANALYSIS OF 25-HYDROXYVITAMIN D IN SERUM USING UPLC/MS/MS

Waters
THE SCIENCE OF WHAT'S POSSIBLE.™

LJ Calton¹, SD Gillingwater¹, GW Hammond¹, DP Cooper¹ & S Wilson²
¹Waters Corporation, Manchester, UK ²Waters, Australia

INTRODUCTION

Several recent studies have shown that vitamin D deficiency is common in adults and children around the world. In addition to the well known effects of vitamin D deficiency, such as calcium malabsorption, there is growing evidence that the risk for other conditions (e.g. cancers^{1,2}) may be increased. The measurement of 25-hydroxyvitamin D [25(OH)D] is accepted as the clinical indicator of vitamin D status³ and is important in the diagnosis and treatment of vitamin D deficiency. The major issue with immunoassays is that they cannot differentiate between the two forms of 25(OH)D: 25(OH)D₂ & 25(OH)D₃, and instead, rely on the cross-reactivity of the antibody to measure a total 25(OH)D concentration. If that cross-reactivity is less than 100% then vitamin D₂ therapy may not be monitored effectively.⁴ The aim of this study was to develop a quantitative method for 25(OH)D₂ and 25(OH)D₃ in serum to prevent the misdiagnosis of vitamin D deficiency in patients.



Figure 1. System configuration of Waters ACQUITY UPLC / TQD

METHODS

A Waters® ACQUITY® Tandem Quadrupole Detector (TQD) coupled to an ACQUITY UPLC® (Waters Corporation, Manchester, UK) was used for all analyses (Figure 1). The 25(OH)D compounds were separated from endogenous interferences using an ACQUITY UPLC BEH C8 Column 2.1 x 50 mm, 1.7 µm employing a gradient elution profile, 73-98% B in 1.5min following a 2min initial hold at a flow rate of 0.4ml/min, where mobile phase A and B are 2mM ammonium acetate+0.1% formic acid in water and methanol respectively.

The instrument was operated in positive electrospray ionisation mode using MassLynx™ 4.1 software with auto data processing by the QuantLynx™ Application Manager. Specific Multiple Reaction Monitoring (MRM) experiments for each compound were created as shown in Table 1.

A single calibrator and bi-level QC's (Chromsystems, Munich, Germany) were prepared as per the manufacturer's instructions. A low QC was prepared by pooling human serum and adding a known concentration of 25(OH)D₂ and 25(OH)D₃. The final concentrations of the low, medium and high QC samples were 19, 27 and 84ng/mL for 25(OH)D₂ and 13, 29 and 89ng/mL for 25(OH)D₃ respectively.

Compound	MRM	Dwell (secs)	Core Voltage (V)	Collision Energy (eV)
25(OH)D ₂	401.35 >150.1	0.05	24	28
25(OH)D ₃ *	401.35 >260.3	0.05	24	10
4 _α -25(OH)D ₃	407.35 >150.3	0.05	24	28
25(OH)D ₂	413.35 >81.1	0.05	24	22
25(OH)D ₃ *	413.35 >265.3	0.05	24	10

Table 1. The tuning parameters used when monitoring for 25(OH)D₂ and 25(OH)D₃ and the internal standard. *denotes optional qualifier ion

To assess linearity, calibrators were prepared in mammalian serum over the concentration range 2.5-100ng/mL for 25(OH)D₂ and 25(OH)D₃.

The samples were prepared using a liquid-liquid-extraction protocol that involves the addition of internal standard (250ng/mL hexa-deuterated 25(OH)D₃, Synthetica AS, in 80% MeOH/20% IPA), ZnSO₄, MeOH and Hexane to 350µL of serum. Following centrifugation for 5 mins at 13,000rpm, the hexane layer was removed and placed into Waters maximum recovery vials and evaporated to dryness under nitrogen at 50°C. The samples were reconstituted in 75µL of 70% methanol in water and 20µL was injected.

RESULTS

Accuracy

The accuracy of the assay was determined by the analysis of external quality control samples from DEQAS (www.deqas.org). The Chromsystems single point calibrator was used and a calibration line constructed through zero to calculate the DEQAS sample concentrations. Passing-Babik linear regression was used to compare the Waters 25(OH)D₃ results with the DEQAS LC/MS method mean. All results were within ±11.5% deviation of the expected value (Figure 2).

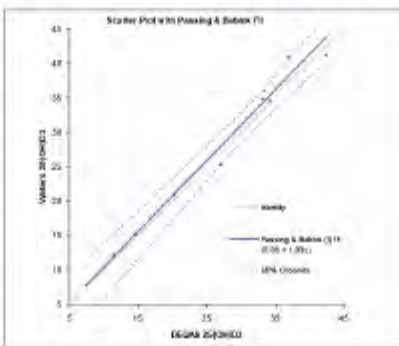


Figure 2. Passing-Babik linear regression analysis comparing the Waters 25(OH)D₃ results to DEQAS LC/MS method mean

Linearity

The coefficient of determination (R²) for 25(OH)D₃ was >0.999 and the calculated concentrations for the calibrators were all within ±4% of the assigned values. The coefficient of determination (R²) for 25(OH)D₂ was >0.997 and the calculated concentrations for the calibrators were all within ±10% of the assigned values.

Precision

The intra-assay precision was determined by extracting and analysing five replicates of the low, medium and high QC samples. The coefficient of variation (CV) for 25(OH)D over the three levels were calculated. The inter-assay precision was determined over five consecutive days using the low, medium and high QC samples. The results are shown in Table 2.

	Low QC		Medium QC		High QC	
	25(OH)D ₂	25(OH)D ₃	25(OH)D ₂	25(OH)D ₃	25(OH)D ₂	25(OH)D ₃
Intra-assay % CV	5.4	7.5	8.0	3.8	3.1	6.2
Inter-assay % CV	8.7	6.3	9.2	5.5	7.2	5.9

Table 2. Summary of the intra and inter-assay precision of the assay

DISCUSSION

A method for the UPLC/MS/MS analysis of 25(OH)D₂ and 25(OH)D₃ in serum has been developed. The methodology involves a simple liquid-liquid extraction of the analytes from serum and the MRM detection of each analyte using two transitions. Quantifier and qualifier ion ratios were monitored to ensure lack of interference⁵. The assay demonstrates good sensitivity with acceptable intra and inter-day precision. Using this methodology it is feasible to manually process and analyse up to 100 samples per day.

CONCLUSION

- A method for the independent quantification of 25(OH)D₂ and 25(OH)D₃ in serum has been developed with good linearity, sensitivity and precision.
- UPLC/MS/MS offers significant advantages over the traditional HPLC/UV methodology through reduced sample volume, increased sensitivity, specificity and speed.
- UPLC/MS/MS allows for the accurate and reliable measurement of 25(OH)D₂ and 25(OH)D₃ in serum to prevent the misdiagnosis of vitamin D deficiency in patients who are receiving vitamin D₂ supplementation.

References

1. Gannan HO, Gannan CH, Gannan PC, Ward WB, Miller M, Lujan M, et al. Optimal vitamin D intake for colorectal cancer prevention: a quantitative meta-analysis. *Am J Prev Med* 2007;32:210-6.
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5. Cook MS. Spectrometry in the clinical laboratory: general principles and guidelines. CSHL, 2001.

Method comparisons

25-Hydroxyvitamin D - Which Assay?

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Introduction

Vitamin D plays an important role in calcium homeostasis. Deficiency is associated with defects in bone mineralisation, and may predispose to a range of progressive and immune diseases (1). Low levels are prevalent in many European populations and dietary supplementation is a major strategy to reduce fracture risk, particularly in the elderly (2). Accurate assessment of Vitamin D status is essential. Both 25-hydroxyvitamin D (25(OH)D) and 1,25-dihydroxyvitamin D (1,25(OH)₂D) are present in serum and reflect treatment. A number of different methods of 25(OH)D measurement are available. The most commonly used method is immunoassay. However, immunoassays are subject to a number of limitations, including cross-reactivity with other vitamin D metabolites and the presence of interfering substances. The gold standard method for 25(OH)D measurement is liquid chromatography-tandem mass spectrometry (LC-MS/MS) (3).

Analysis of 25(OH)D is complicated by the requirement to detect both 25-hydroxyvitamin D₃ (25(OH)D₃) and 25-hydroxyvitamin D₂ (25(OH)D₂). Poor agreement between 25(OH)D immunoassays reported in recent quality assurance programs (4) has raised concerns about their ability to accurately assess Vitamin D status, particularly in patients taking Vitamin D. This call is answered by use of an agreed reference method with which to assess performance. Recently, the Institute of Reference and Diagnostic Chemistry (IRDC) and the International Federation of Clinical Chemistry (IFCC) have agreed to use LC-MS/MS as a reference method for 25(OH)D analysis.

AIM

To use LC-MS/MS as a reference method to:
1) investigate the presence of 25(OH)D₂ in routine patient sera; and
2) assess the accuracy of five 25(OH)D immunoassays in routine containing or not significant amounts of the metabolite.

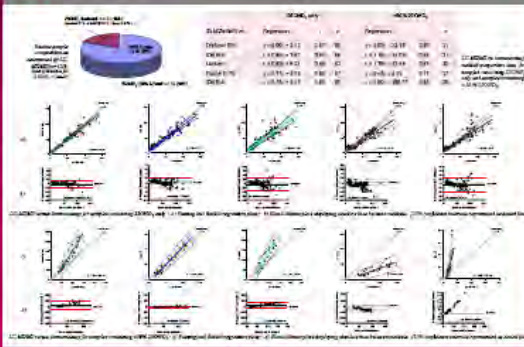
Methods

120 unlabelled routine 25(OH)D serum samples were collected over a 12 week period in 2006. Of these, 107 were obtained from The Royal Children's Hospital (RCH) (Paediatric Service) and 13 were from the Royal Children's Hospital (RCH) (Paediatric Service) from a related general adult population. An additional 121 samples were collected from nine clinical trials, subjected to high doses of Vitamin D to enable sample collection and analysis. Approval for the use of all samples was granted by both the RCH Ethics and Human Research Committee and the RCH Human Research Ethics Committee. 25(OH)D₂, 25(OH)D₃ and total 25(OH)D concentrations were determined using LC-MS/MS (5).

25(OH)D was measured using the following five immunoassay systems:
• DiaSorin Liaison (immunoassay) (LIA)
• IDS iCHEM RIA
• DiaSorin Liaison² Automated Chemiluminescent Immunoassay (LIA²)
• Roche Elecsys² Automated Chemiluminescent Immunoassay (CIA)
• IDS OPTIA² Automated Chemiluminescent Immunoassay (CIA)

All samples were stored frozen until 2007 in separate aliquots for each assay and subjected to three freeze-thaw cycles to simulate routine analysis (6,7). Results from samples containing 100% 25(OH)D₂ and 100% 25(OH)D₃ were assessed against LC-MS/MS using assays 1-5 (Diagn. Laboratory with Abbott Eutek 200).

Results



Discussion

When samples contained only 25(OH)D₃, all immunoassays demonstrated a similar high negative bias compared to LC-MS/MS, with the lowest bias coming from the IDS RIA in a routine assay. Agreement with LC-MS/MS was much more variable when samples contained significant levels of 25(OH)D₂. The Roche assay greatly overestimated 25(OH)D₂ when not adjusted as the antibody specificity for 25(OH)D₂ is stated as zero in the kit insert. This assay is marketed as a 25(OH)D₃ assay and is unsuitable for monitoring 25(OH)D in patients taking VitD.

The two DiaSorin assays (LIA and Liaison) demonstrated a positive proportion bias in the group which was unexpected as the antibody specificity for 25(OH)D₂ is stated as 100%. Similarly, the IDS RIA showed an unexpectedly good correlation, given an antibody specificity for 25(OH)D₂ of 75%. It is proposed that these assays may detect additional metabolites, such as 24-hydroxyvitamin D₂, which have been detected in the serum of subjects taking high doses of VitD (8). It is unclear why the IDS RIA produced such higher results than the DiaSorin, as the 25(OH)D₂ specificity in these kits is stated to be the same.

While most routine clinical samples contained only 25(OH)D₃, 25(OH)D₂ was detected in 18% including that VitD₂ is still seen as a strong supplement and we should remain concerned about the accuracy of routine 25(OH)D assays in the presence of this metabolite.

Conclusions

All assays demonstrated a similar, slight negative bias compared with LC-MS/MS when samples contained 25(OH)D₃ only. Agreement was more variable in samples containing 25(OH)D₂. The presence of this metabolite in 18% of routine patients indicates the VitD₂ is still seen as a strong supplement and routine 25(OH)D results should be interpreted with caution.

References

1. Holmberg M, et al. (2002) Vitamin D deficiency in the elderly. *Journal of Internal Medicine* 252: 187-198.
2. Holmberg M, et al. (2002) Vitamin D deficiency in the elderly. *Journal of Internal Medicine* 252: 187-198.
3. Holmberg M, et al. (2002) Vitamin D deficiency in the elderly. *Journal of Internal Medicine* 252: 187-198.
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Original Article

Accuracy and clinical implications of seven 25-hydroxyvitamin D methods compared with liquid chromatography-tandem mass spectrometry as a reference

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2009 QAP end of cycle 32 report

Welcome						Program Selection						End Of Cycle Report: Vitamin D3 (25-hydroxycholecalciferol) Cycle: 32						Summary Data					
Vitamin D3 (25-hydroxycholecalciferol) (nmol/L) - Summary Data Endocrine Program Cycle 32																							
Analytical Principle						No. Labs	S.D.	CV	Low 6.0	High 254.0	Instrument						No. Labs	S.D.	CV	Low 6.0	High 254.0		
Isotope Diln Tandem Mass Spec (IDMSMS)						4	5.4	4.45	7.0	242.0	Applied Biosystems API 3200 Q-TRAP						2	4.3	3.4	7.0	246.0		
Radioimmunoassay						5	8.9	9.0	12.0	214.0	HPLC Waters						1	5.3	4.4	6.0	238.0		
Electrochemiluminescence						26	12.5	11.75	44.0	160.0	Applied Biosystems API 4000 Q-TRAP						1	6.4	5.0	8.0	246.0		
HPLC						1	15.5	12.5	3.0	245.0	Nichols Institute Diagnostics Advantage						1	5.2	5.2	38.0	162.0		
Chemiluminescence						24	15.0	15.3	36.0	170.0	Roche Diagnostics Hitachi Modular						1	8.1	8.0	46.0	156.0		
ELISA						4	20.3	19.25	44.0	186.0	Scintillation Counter - Gamma						5	8.9	9.0	12.0	214.0		
Roche Diagnostics Elecsys 1010/2010/cobas e 411						4	11.3	10.9	46.0	159.0	Roche Diagnostics E170/ e 601 (cobas 6000-IA)						21	12.6	12.9	43.0	161.0		
Own Preparation						3	5.5	4.5	8.0	246.0	DiaSorin Liaison						23	15.4	15.4	35.0	170.0		
Roche Diagnostics (Integra)						1	5.4	4.7	44.0	183.0	Spectrophotometer/Plate Reader Spectrophotometer/Plate Reader						3	19.5	20.9	42.0	183.0		
Chromsystems						2	10.4	8.45	5.0	242.0													
Roche Diagnostics (Hitachi)						25	12.6	12.9	44.0	159.0													
DiaSorin						28	14.3	14.75	32.0	172.0													
IDS Ltd						5	19.5	17.6	42.0	183.0													
Reagent						No. Labs	S.D.	CV	Low 6.0	High 254.0													
Calibrator						No. Labs	S.D.	CV	Low 6.0	High 254.0													
Chromsystems						1	6.4	5.0	8.0	246.0													
Inhouse Calibrator						5	9.9	9.5	42.0	165.0													
LC-MS-MS						21	13.6	14.1	42.0	159.0													
UV Quantification						4	17.5	14.7	29.0	202.0													

Your Method Code: I: Electrochemiluminescence 11L : Roche Diagnostics E170/ e 601 (cobas 6000- IA) 21: Roche Diagnostics (Hitachi) C: LC-MS-MS

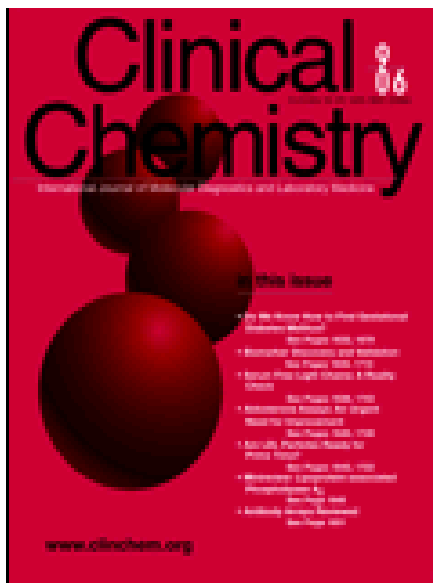
Jan 2010 DEQAS: Vitamin D

25-HYDROXYVITAMIN D3							
		346	347	348	349	350	
HPLC	Mean	29.9	93.4	24.6	47.2	48.8	nmol/L
	SD	8.3	21.6	6.0	9.0	9.3	nmol/L
	n	19	19	19	19	19	
	cv	28	23	25	19	19	%
LCMS	Mean	31.3	94.2	25.1	48.0	50.5	nmol/L
	SD	5.4	14.0	5.1	7.1	11.7	nmol/L
	n	43	43	43	43	43	
	cv	17.2	14.9	20.3	14.8	23.1	%

LC-MS/MS Reference methods

- Various LC-MS/MS methods available
- Variation between these methods
- Roche assay based on the method below
- This was developed in cooperation with Dr. Vogeser (Klinikum Grosshadern)
- Then further optimized at Roche

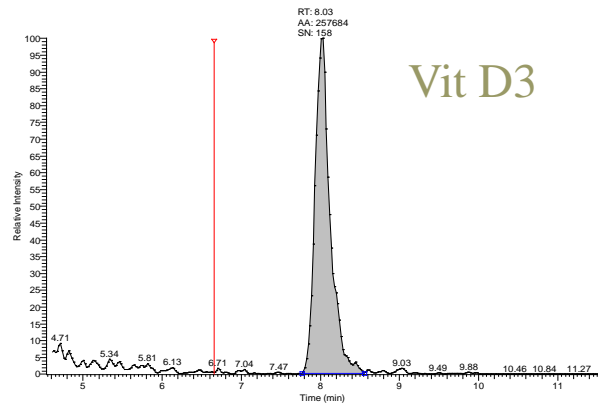
Clinical Chemistry 50, No. 8, 2004



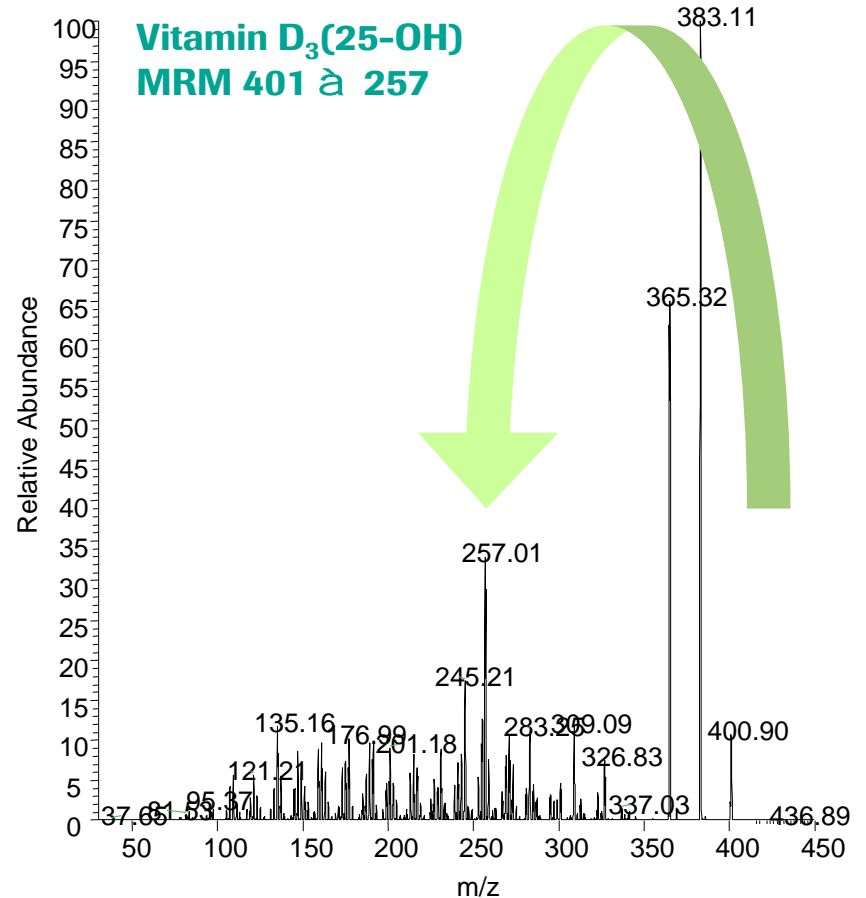
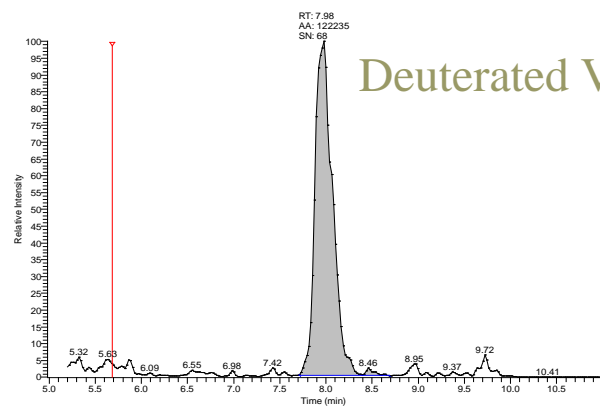
Candidate Reference Method for the Quantification of Circulating 25-Hydroxyvitamin D₃ by Liquid Chromatography–Tandem Mass Spectrometry, Michael Vogeser,^{1*} Apostolos Kyriatsoulis,² Erasmus Huber,² and Ilwe Kobold²
(¹ Institute of Clinical Chemistry, Hospital of the University of Munich, Munich, Germany; ² Roche Diagnostics GmbH, Penzberg, Germany; * address correspondence to this author at: Institute of Clinical Chemistry, Hospital of the University of Munich, D-81366 Munich, Germany; fax 49-89-7095-3240, e-mail Michael.Vogeser@med.uni-muenchen.de)

Vit D3 chromatography + MS/MS

OP_15Jan2009_13 - TIC - SM: 6 RT: 4.53 - 11.53 NL: 1.93E4
F: + cAPCI sid=5.00 SRM ms2 401.300@33.00 [159.150-159.250, 257.150-257.250]



OP_15Jan2009_13 - TIC - SM: 5 RT: 4.98 - 10.98 NL: 9.15E3
F: + cAPCI sid=5.00 SRM ms2 407.300@33.00 [159.150-159.250, 263.150-263.250]



Vitamin D3 (25-OH) elutes at approx. 8 min.

total gradient time: 20 min. incl. extensive column cleaning

Current challenges in vitamin D standardization

- *There is considerable variability in reference methods* Lack of a „real“ vitamin D standard reference material which can be used for immunoassays
- Variability in methods for reference standardization (methodological risks, influence of chromatographic resolution)
- No “real“ reference values existing

NIST human serum SRM

SRM	Description	Certified Constituents	Reference	Form	No. of Levels
909b	Human Serum	Calcium, Chloride, Cholesterol, Creatinine, Lithium, Magnesium, Potassium, Sodium, Total Glycerides, Triglycerides, Urea, and Uric Acid	Bilirubin	Lyophilized	2
1951b	Lipids in Frozen Human Serum	Total Cholesterol, Total Glycerides, Triglycerides		Frozen	2
956b	Electrolytes in Frozen Human Serum	Total Ca, Li, Mg, K, Na	Ionized Ca	Frozen	3
965a	Glucose in Frozen Human Serum	Glucose		Frozen	3
967	Creatinine in Frozen Serum	Creatinine		Frozen	2
970	Ascorbic Acid in Frozen Human Serum	Total Ascorbic Acid		Frozen	2
1952a	Cholesterol in Human Serum (Freeze-dried)	Cholesterol		Lyophilized	3
968c	Fat-Soluble Vitamins, Carotenoids, and Cholesterol in Human Serum	Vitamins (4), Cholesterol, Carotenoids (4)	Carotenoids (8), Vitamin D	Lyophilized	2
1589a	PCBs, Pesticides, and Dioxins/Furans in Human Serum	PCB Congeners (16), Chlorinated Pesticides (5), Total Cholesterol	PCB Congeners (9), Chlorinated Pesticides (5), Total Cholesterol, Triglycerides, "Free" Cholesterol, Phospholipids	Lyophilized	1
1599	Anticonvulsant Drug Level Assay (valproic acid and carbamazepine)	valproic acid carbamazepine		Lyophilized	1
900	Antiepilepsy Drug Level Assay	Antiepileptics (4)		Lyophilized	3
1955	Homocysteine and Folate in Human Serum	Homocysteine 5-Methyltetrahydrofolic acid	Total Folate, Folic Acid	Frozen	3



Vit D
Released 2009
SRM 972

Allows for a
common
primary
calibrator

Now 968d

Current challenges in vitamin D standardization

- *Limitations of NIST controls SRM 972:*
 - n Level 1: native human serum
 - n Level 2: level 1 diluted with horse serum
 - n Level 3: human serum spiked with vitamin D2 (25-OH)
 - n Level 4: human serum spiked with vitamin D3 (25-OH) and 3-epi 25(OH)
- *In vitro anomaly affecting immunoassays*
 - n Exogeneously added vitamin D does not distribute to the vitamin D binding protein (VDBP) as it occurs as in vivo
 - n Exogeneously added material binds to other moieties than the VDBP
- *à failure of quantitative recovery in immunoassays*
- *Is there a way out of this dilemma ?*

NIST SRM 972 standard is detectable by the Roche LC-MS/MS

NIST Level	Target conc. Vit. D3 (25-OH)	Target conc. 3-epi-Vit.D3 (25-OH)	Target conc. Vit.D2 (25-OH)	NIST total Vitamin D	Conc. found with Roche LC-MS/MS		Total Vitamin D Roche LC-MS/MS
					Vit. D3 (25-OH)	Vit. D2(25-OH)	
Level 1	23.9 +/- 0.8	1.39 +/- 0.04	0.60 +/- 0.20	25.9	24.8	1.5	26.3
Level 2	12.3 +/- 0.8	0.76 +/- 0.02	1.71 +/- 0.08	14.8	14.6	1.2	15.8
Level 3	18.5 +/- 1.1	1.06 +/- 0.03	26.4 +/- 2.0	46.0	20.5	25.2	45.7
Level 4	33.0 +/- 0.8	37.7 +/- 1.2	2.4 +/- 0.21	73.1	72.1	3.1	75.2

How safe are LC-MS/MS data ?

The Quest Story

08. Jan 2009 - New York Times:

- **“Quest acknowledges errors in vitamin D tests”**
- The nation's largest medical laboratory company provided possibly **erroneous results to thousands of people** who had their vitamin D levels tested in the last two years, the company has acknowledged.
- Quest's problems with the vitamin D analysis arose after it shifted in 2006 and 2007 to a new test of its own design, replacing an older F.D.A.-approved test.
- The new test promised to be more accurate and offer more detailed information, Quest executives said. But the test relied on a **sophisticated instrument called a mass spectrometer**, which can be tricky to use, especially for high-volume testing.

Specifications for trueness and precision of a reference measurement system for serum/plasma 25-hydroxy vitamin D analysis

Clinica Chimica Acta, 2009; 408: 8-13

Dietmar Stöckl, Patrick M. Sluss and Linda M. Thienpont

Abstract

Background

The divergence in analytical quality of serum/plasma 25-hydroxy-vitamin D analysis calls for defining specifications for a reference measurement system.

Methods

Fundamentally, in a reference measurement system, there should be a relationship between the analytical specifications for higher- (reference) and lower-order (routine) measurements. Therefore, when setting specifications, we started with limits for routine imprecision (CV_{rou}) and bias (B_{rou}) using 4 models: (1) the misclassifications in diagnosis, (2) biological variation data (reference interval (RI) and monitoring), (3) expert recommendations, and (4) state-of-the-art performance. Then, we used the derived goals to tailor those for reference measurements and certified reference materials (CRMs) for calibration by setting the limits for CV_{ref} at $0.5 CV_{rou}$, B_{ref} at $0.33 B_{rou}$, max. uncertainty (U_{max}) at $0.33 B_{ref}$

Results

The established specifications ranged between $CV_{rou} \leq 22\%$, $B_{rou} \leq 10\%$, $CV_{ref} \leq 11\%$, $B_{ref} \leq 3.3\%$, $U_{max} 1.1\%$ (model 3) and $CV_{rou} \leq 4\%$, $B_{rou} \leq 2.6\%$, $CV_{ref} \leq 2\%$, $B_{ref} \leq 0.9\%$, $U_{max} 0.3\%$ (model 2, monitoring).

Conclusions

Model 2 (monitoring) gave the most stringent goals, model 3, the most liberal ones. Accounting for state-of-the-art performance and certification capabilities, we used model 2 (RI) to recommend achievable goals: for routine testing, $CV_{rou} \leq 10\%$, $B_{rou} \leq 5\%$, for reference measurements, $CV_{ref} \leq 5\%$, $B_{ref} \leq 1.7\%$, and for CRMs, $U_{max} 0.6\%$.

Keywords: Serum/plasma 25-hydroxyvitamin D2; Serum/plasma 25-hydroxyvitamin D3; Quality goals; Bias; Imprecision; Uncertainty

Stockl *et al*: Approaches for quality

- Four different approaches
- Followed the 1999 Stockholm consensus conference guidelines on quality specifications
 1. Clinical Interpretation: Analyses the impact of bias and analytical imprecision on interpretation of results based on clinical decision limits
 2. Biological Variation: Relates analytical performance to the intra- and inter-individual biological variation of vitamin D
 3. Expert Opinion: Considered performance goals set by expert opinion from external quality assurance programs
 4. State of the art: Evaluated the literature on currently used measurement procedures for vitamin D analysis with stated recovery and imprecision data.

Stockl *et al.*: What they found

- Routine measurement systems Expert opinion gave the most liberal goals i.e. 5x the CV's for the biological variation goals
- Expert opinion CV 22% Bias 10%
- Biological Variation CV 4% Bias 2.6%
- Survey of 14 "state of the art" studies of methods
 - **only one method** was close to achieving the performance required by the Biological Variation model i.e. the most stringent model.

Stockl *et al*: Recommendations

- Using the biologic variation approach i.e. Gowan model
- Routine Testing
 - n CV \leq 10%
 - n Bias \leq 5%
- Reference Measurements
 - n CV \leq 5%
 - n Bias \leq 1.7%
- Westgard: “The requirements are challenging, but Stock *et al* believe the laboratory community is up to the challenge”



Moves for Harmonisation

- n LC-MS/MS for standardization of Vitamin D assays needs
 - o high analytical investment
 - o critical interpretation of data (especially in HPLC method validation)
 - o accurate and reliable methods

- n LC-MS/MS methods for standardization can only be compared if specific mass transitions are used

Summary

- Vitamin D: How good are our assays?
 - n Clinical understanding of the importance of vitamin D has increased in recent years
 - n Both automated immunoassays and chromatography MS methods have developed
 - n Discussion surrounds acceptable performance of vitamin D assays
 - n Harmonisation is the next objective